



## **Developing body-on-chip technology for the reduction and replacement of small animals in early drug discovery PK/PD assessments**

**Liam Carr**

University of Edinburgh

Liam.Carr@ed.ac.uk

Disclosure: This presentation covers protected intellectual property, UK patent no PG450503GB

# Overview

- Background
- Hypothesis and aims
- Designing and testing a novel device
- Optimising co-culture
- Kinetic studies
- Future work

# Why?

- 12 years, \$1.3bn per drug
- 25% preclinical success rate (n= 449)
- 7.6% likelihood of approval (n= 3496)

Wouters, et al. *JAMA*, 323(9), 844–853. (2020). <https://doi.org/10.1001/jama.2020.1166>

Takebe et al. *Clinical and translational science*, 11(6), 597–606. (2018). <https://doi.org/10.1111/cts.12577>

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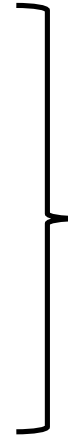
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ALURES – ANIMAL USE REPORTING - EU SYSTEM - SECTION 2. 2020, European Commission.

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Clear need for better early predictors of *in vivo* success

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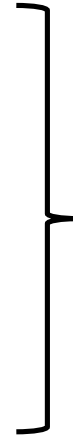
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Clear need for better early predictors of *in vivo* success

- Animal testing of cosmetic products/ingredients banned in EU since 2013
- Push to develop *in vitro*, animal free systems for use in cosmetic product and ingredient safety risk assessments

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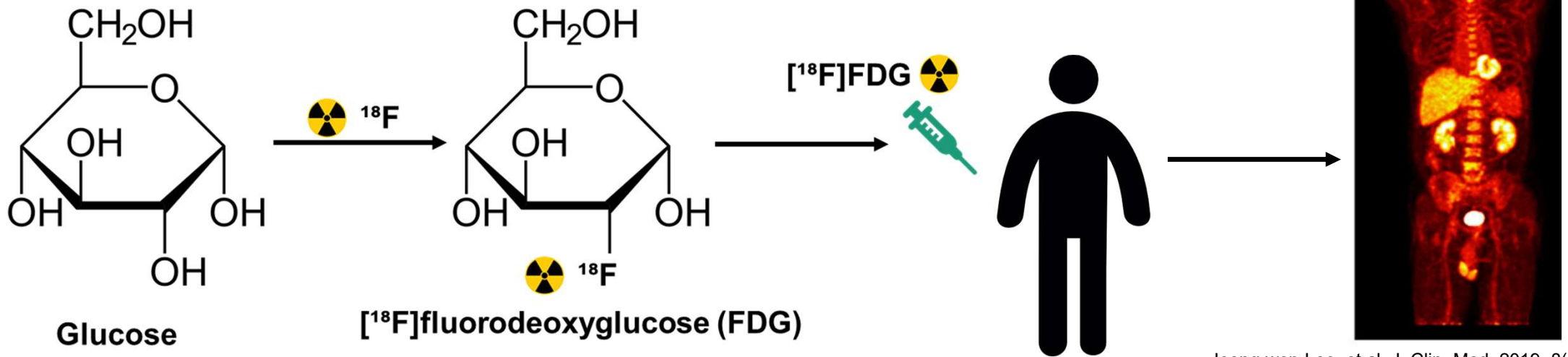
# Positron emission tomography (PET) - what & why?

- High resolution imaging technique utilising a radiotracer
- Short half-life isotopes  $^{18}\text{F}$  (~109min),  $^{68}\text{Ga}$  (~68min), and  $^{11}\text{C}$  (~20min)
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Jeong won Lee, et al. J. Clin. Med. 2019, 8(8), 1169; <https://doi.org/10.3390/jcm8081169>

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# Hypothesis

Body-on-chip platforms capable of circulating drug loaded media across multiple organ compartments can provide PK/PD predictions consistent with that of gold standard *in vivo* human PET data for the same drug.

# Aims

- Optimise the use of a body-on-chip platform such that it is capable of circulating drug-loaded media across multiple “organ” compartments arranged to mimic human physiology.
- Use optimised device to sample “organ” drug concentrations at multiple time points for kinetic modelling
- Compare kinetic parameters to *in vivo* outcomes in human PET studies of the same compound

# Aims

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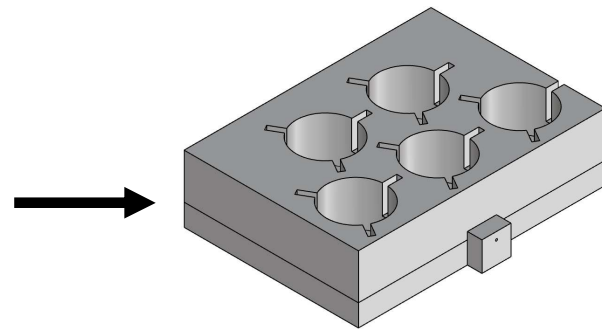
**Brain** = human neurons (SH-SY5Y)

**Lung** = human primary bronchial epithelial cells

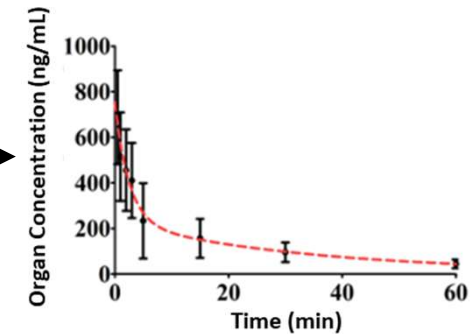
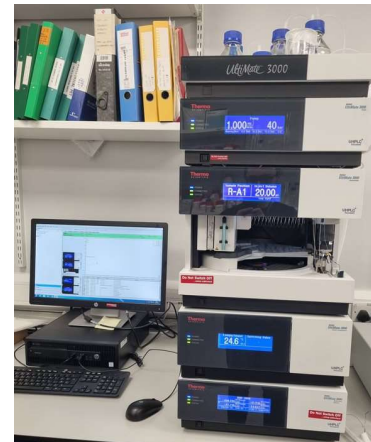
**Liver** = hepatocyte cell line (HepG2)

**Heart** = human primary cardiomyocytes

**Kidney** = Immortalised RPTECs (SA7K)



**Docetaxel/[<sup>18</sup>F]FDG**



Stadulytė *et al.* Journal of chromatography. B. 1118-1119, p33-39; (2019). DOI: <https://doi.org/10.1016/j.jchromb.2019.04.026>

# Overview

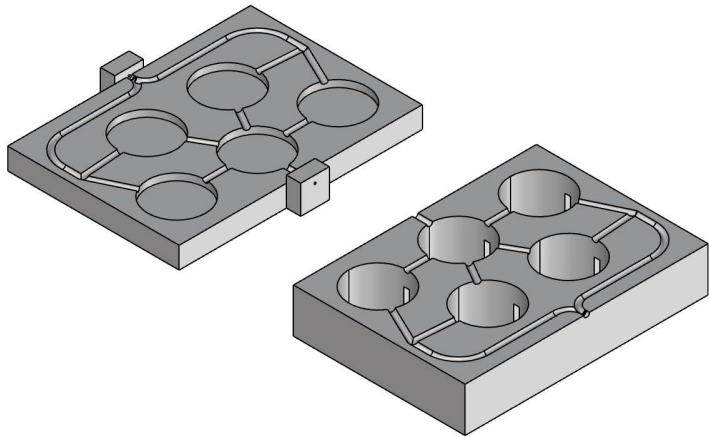
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# Design & test body-on-chip device



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*Edinburgh College of Art*

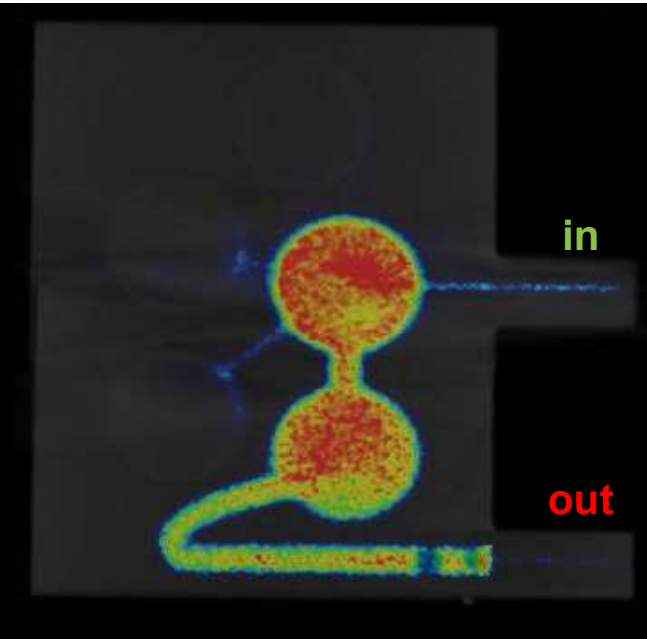
↓  
**Produced 3D  
printed prototypes**



**[<sup>18</sup>F]FDG/[<sup>18</sup>F]NaF PET  
scans to assess flow**



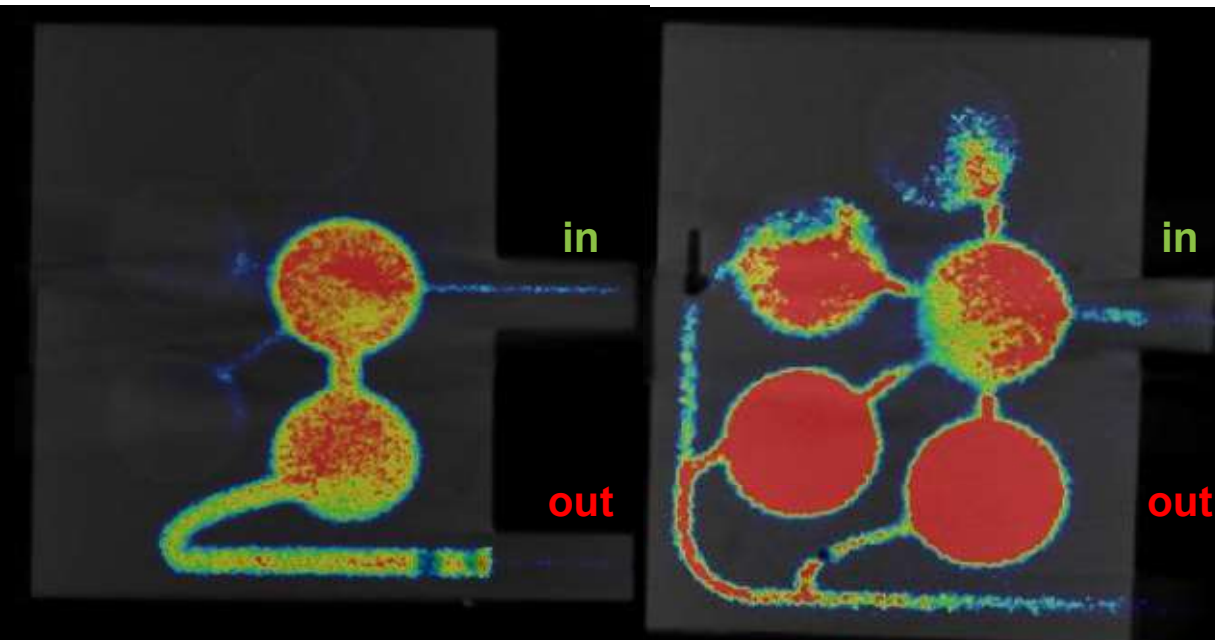
# Design & test body-on-chip device



Capillaries scaled to *in vivo* blood flow:organ volume ratio



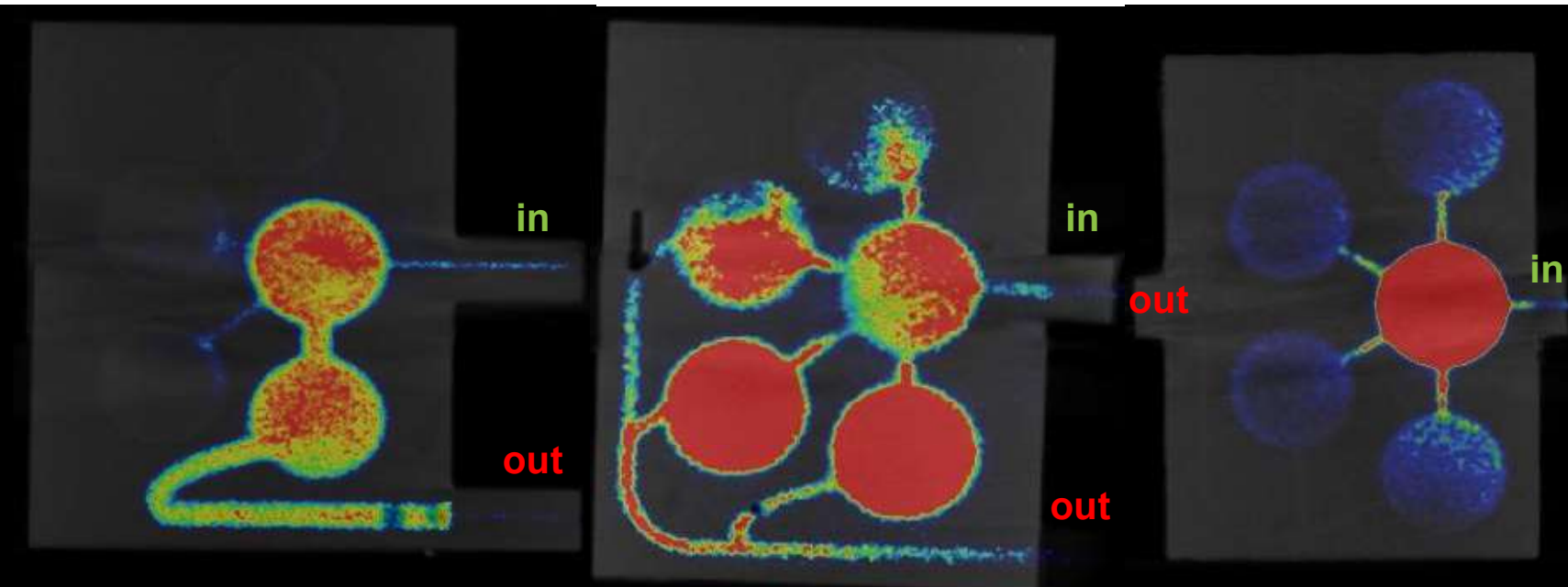
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Capillaries scaled to *in vivo* blood flow:organ volume ratio

Capillaries set to same size (2mm)

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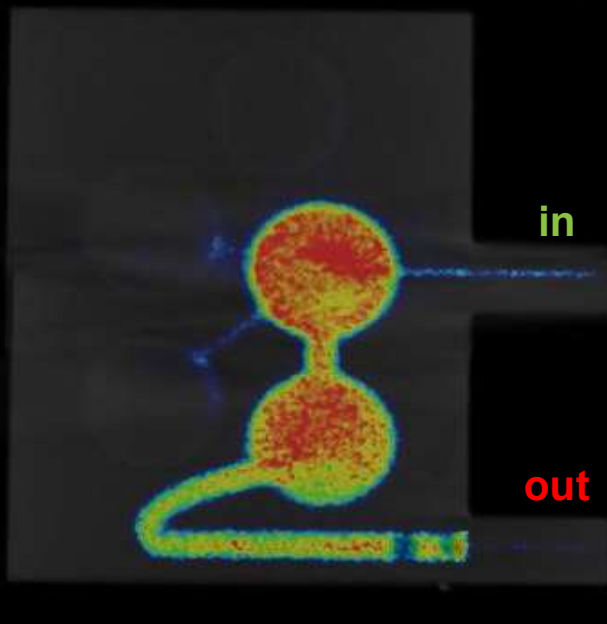


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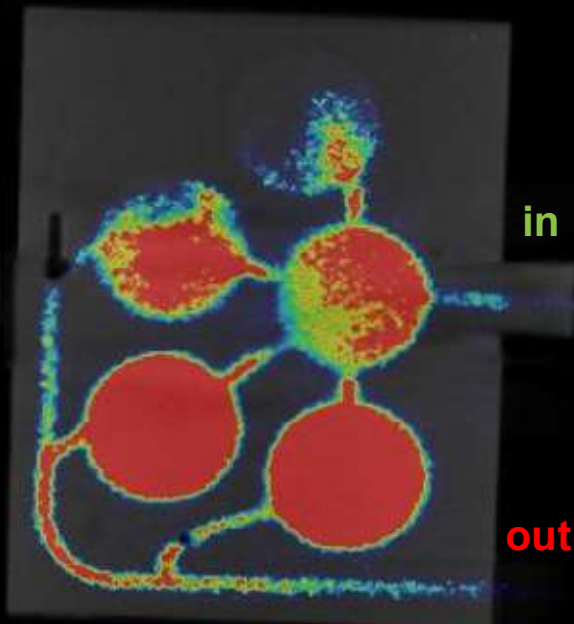
Capillaries set to same size (2mm)

Capillaries same size + completely symmetrical

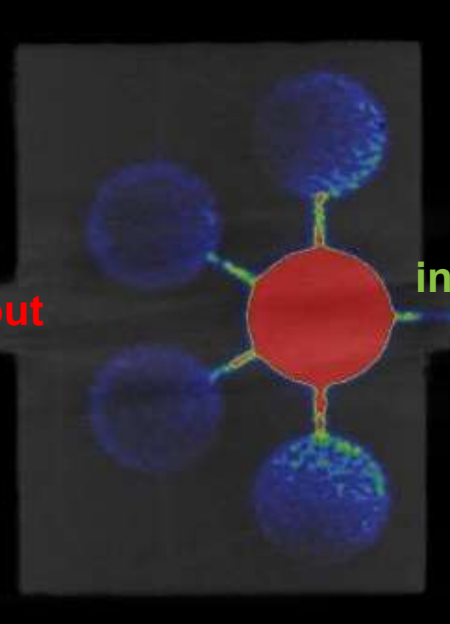
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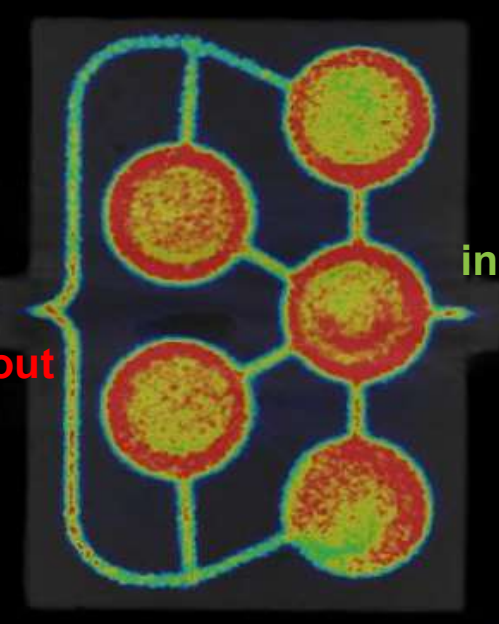
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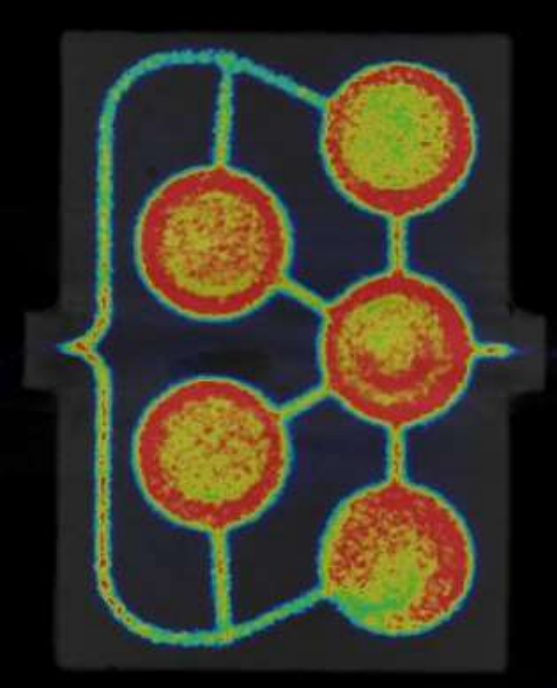
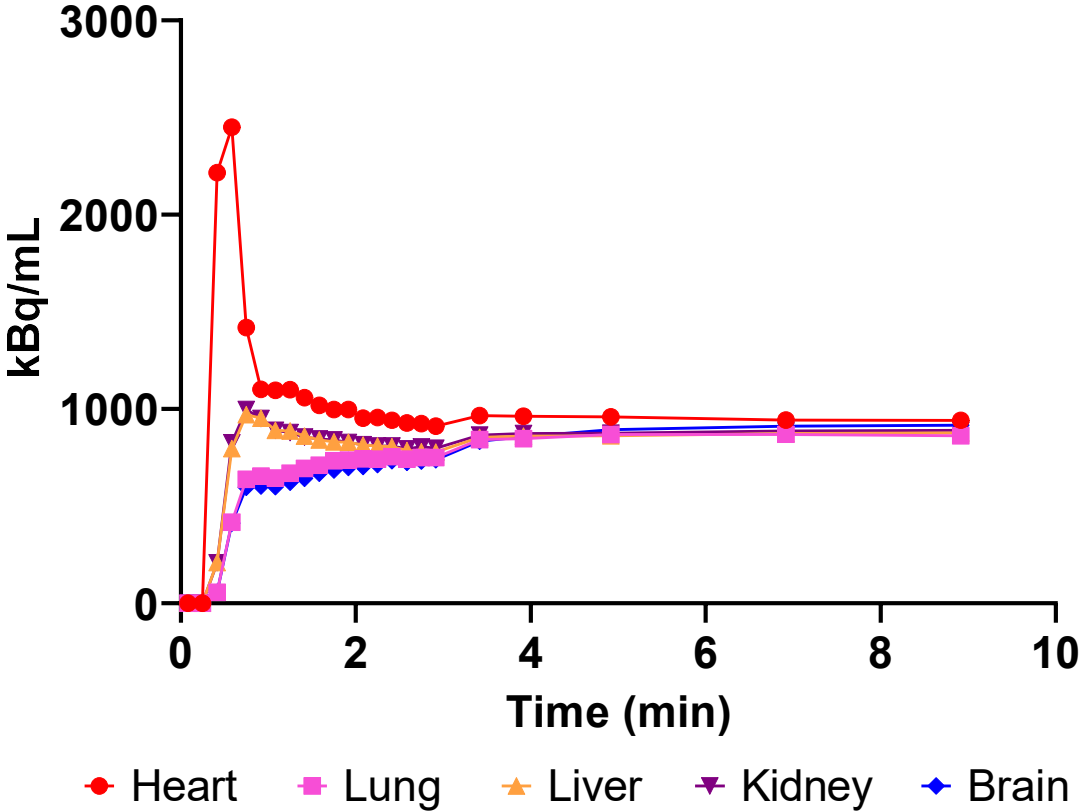


Capillaries same size + completely symmetrical



Capillaries same size + completely symmetrical, with optimised flow rate

# Design & test body-on-chip device



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# Cell culture media optimisation

**Brain** = human neurons  
(SH-SY5Y)

**Lung** = human primary  
bronchial epithelial cells

**Liver** = hepatocyte cell  
line (HepG2)

**Heart** = human primary  
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**Kidney**= Immortalised  
RPTECs (SA7K)

## Common medium?

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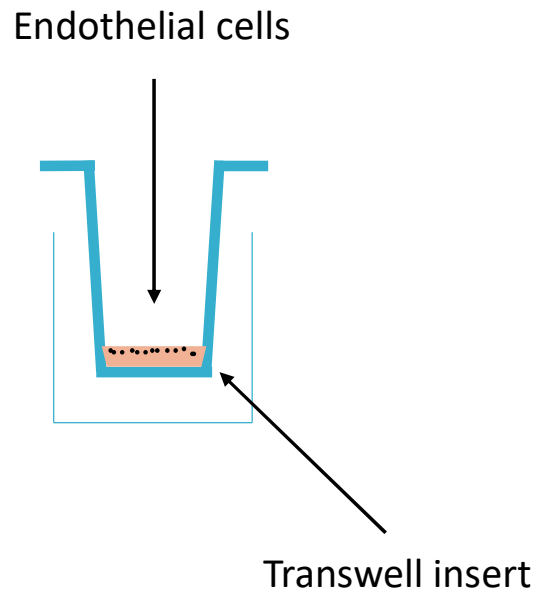
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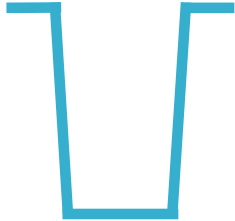
Separation of compartments

# Separation of compartments via endothelial barrier

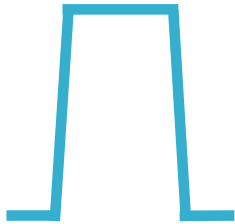




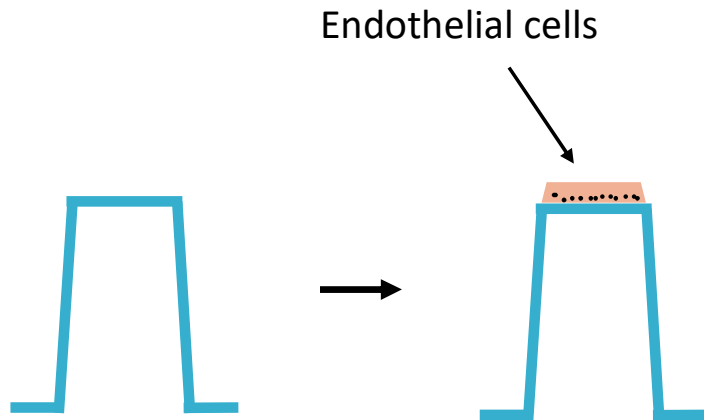
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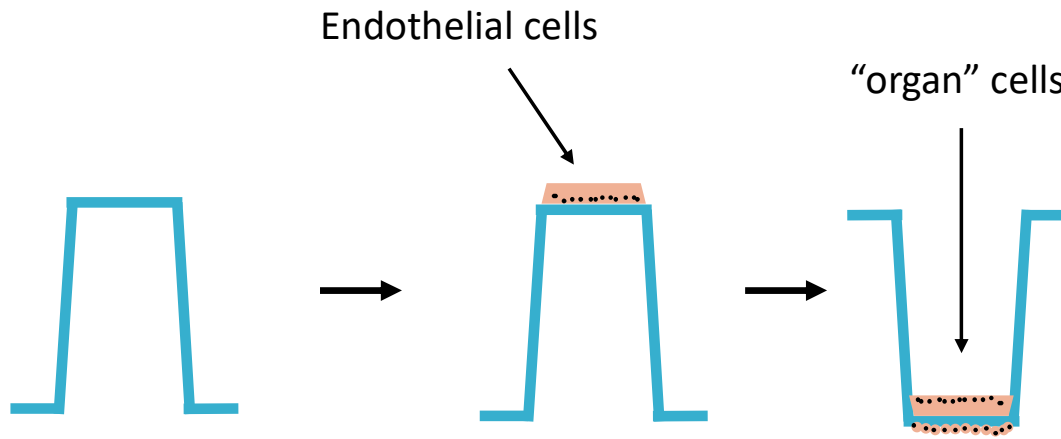
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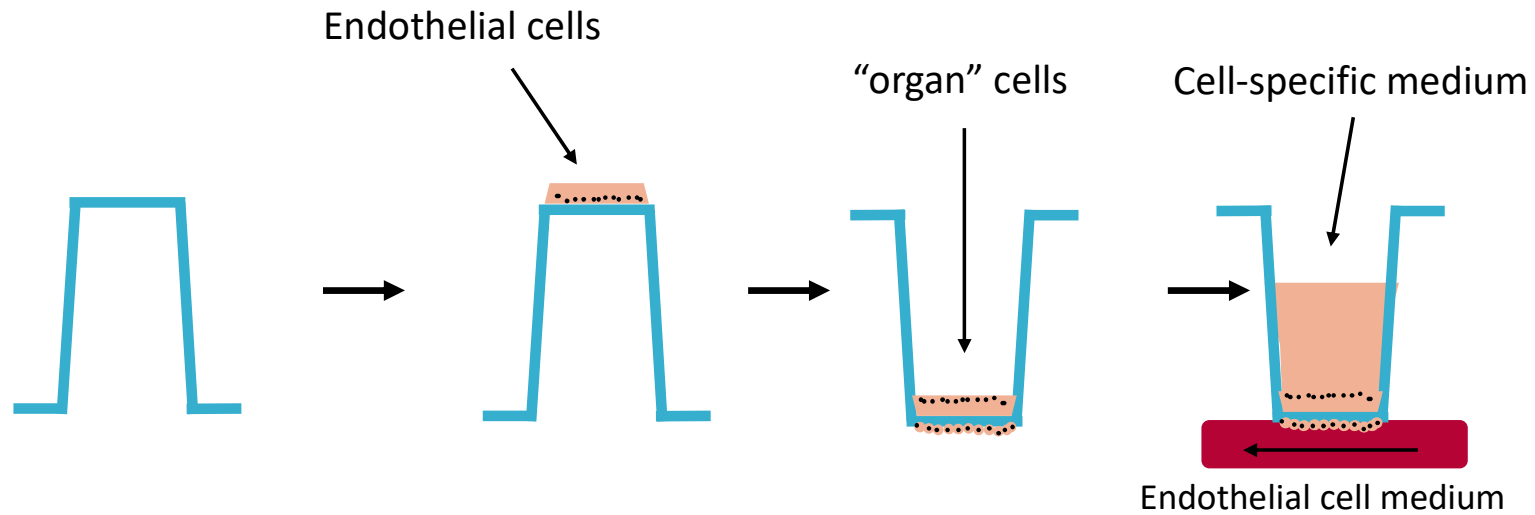
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# Modelling definitions

**$K_i$**  = the rate of influx for a model using irreversible binding (Patlak model for FDG)

**$V_T$**  = Total volume of distribution

**SUV** = standardised uptake value, calculated as concentration in tissue normalised to injected dose and body weight

**SUVmean** = the average SUV across a tissue/organ of interest

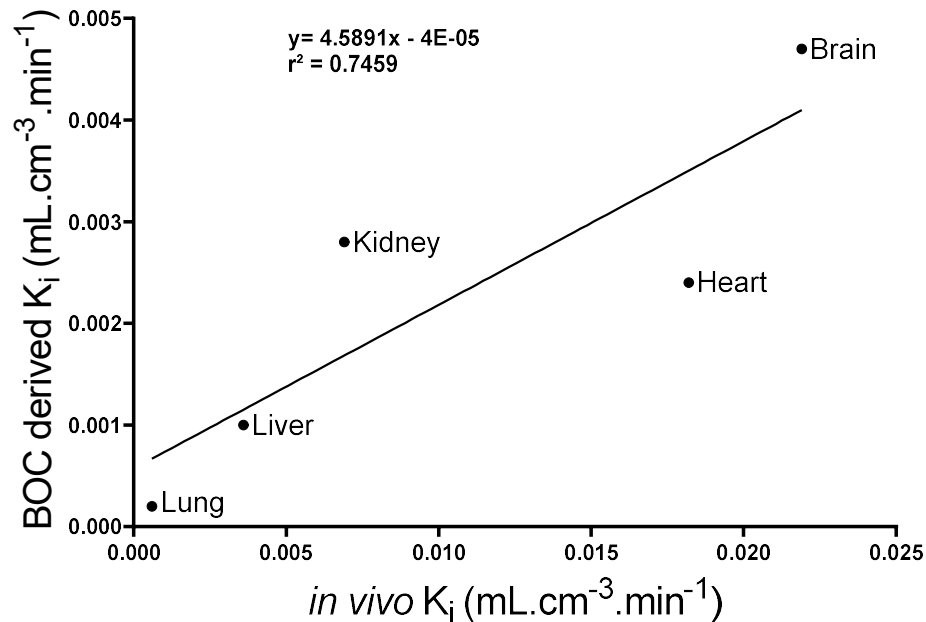
# ***In vitro* FDG $K_i$ significantly correlates with *in vivo* SUVmean**

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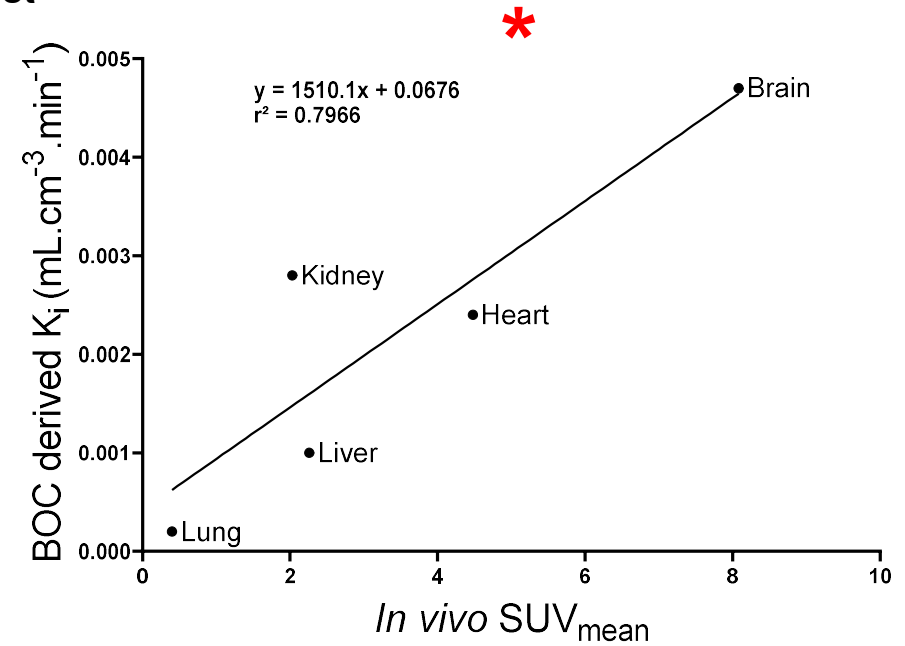
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P= 0.0594, Pearson's correlation, n=1



P= 0.0416, Pearson's correlation, n=1



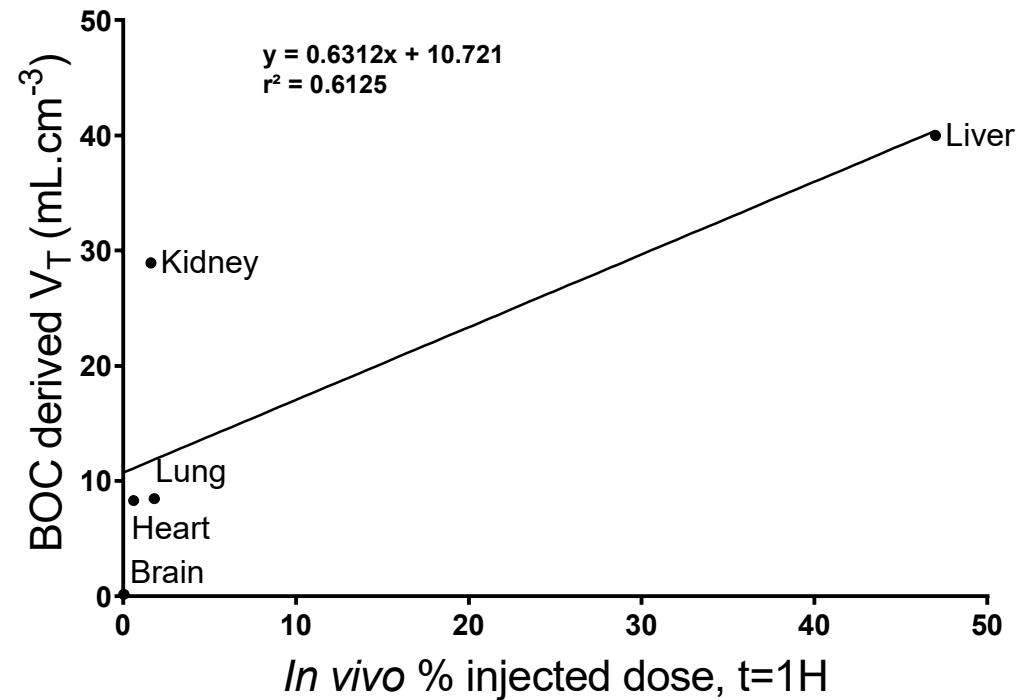
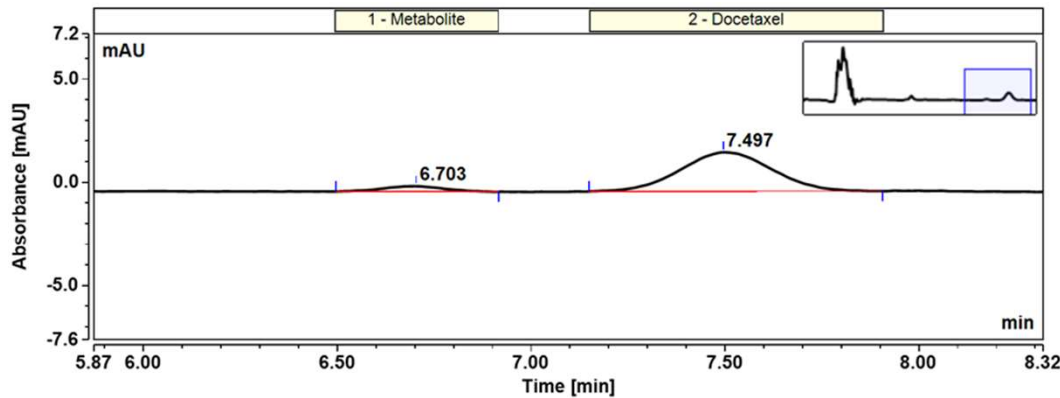
# Device allows for quantification of docetaxel and metabolites

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P= 0.1176, Pearson's correlation, n=1

# Conclusions

- The novel device is capable of housing 5 transwell inserts with even flow through all compartments
- Transwell dual seeding method allows for fluid separation of all compartments without the need for a common medium
- The device can be used to assess rate of influx into tissue, with potential for more accurate predictions of kinetic parameters upon further development
- The device allows for the detection of metabolites as well as assessment of their distribution
- There is clear bias in the elimination compartments (kidney/liver)

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- Slowly increase complexity of the organ compartments
- Incorporate oral absorption via intestinal compartment

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**Medical  
Research  
Council**

## Supervisors:

Dr. Adriana Tavares  
Dr. Mark MacAskill  
Prof. Paddy Hadoke



**Edinburgh Imaging**  
[www.ed.ac.uk/edinburgh-imaging](http://www.ed.ac.uk/edinburgh-imaging)



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## Special thanks

Carlos Alcaide Corral, EPI  
Richard Collins, ECA  
Anne Grant, CRIC



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INNOVATIONS

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Emulating Human Biology

 **SINAPSE**

Scottish Imaging Network: A Platform for Scientific Excellence

# Thank you!

# HPLC LOQ - docetaxel

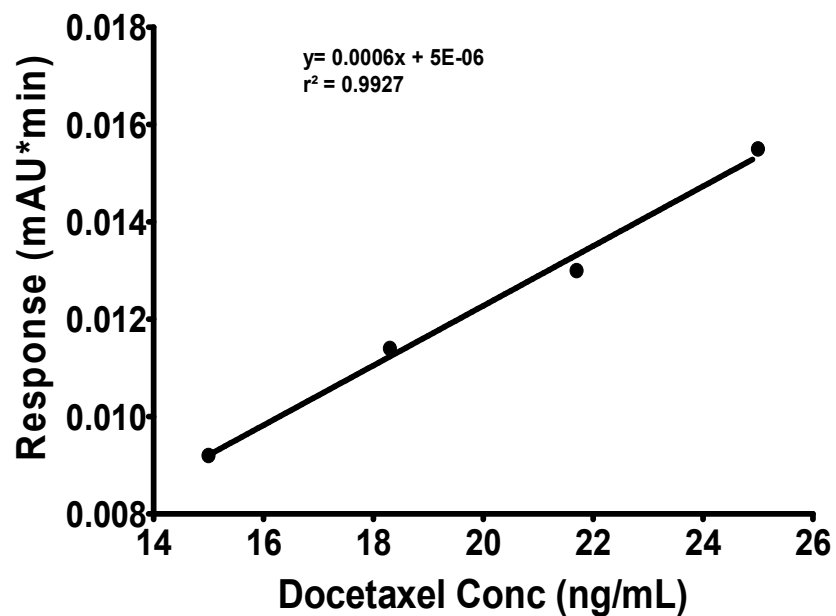
The HPLC LOQ refers to the lowest amount of a compound that can be accurately detected **AND** quantified reliably and accurately. This is calculated as follows:

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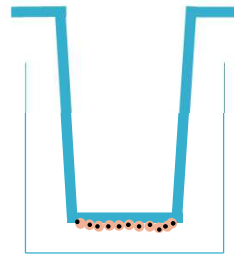
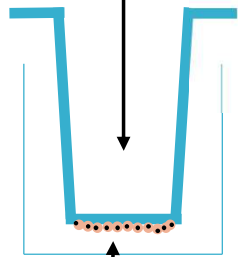
**12.3ng/mL**

# Separation of compartments via endothelial barrier

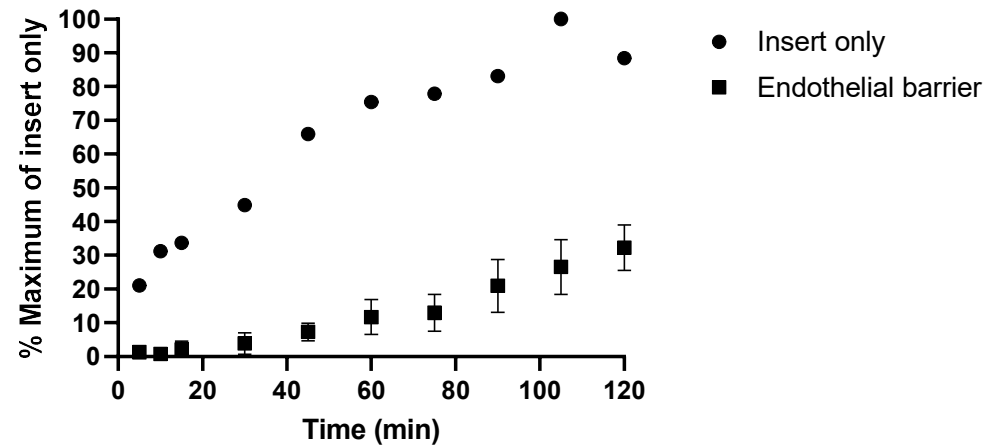
Take small samples over time to measure % Evans blue crossing the barrier



Endothelial medium



Assess using a microplate reader at 610nM



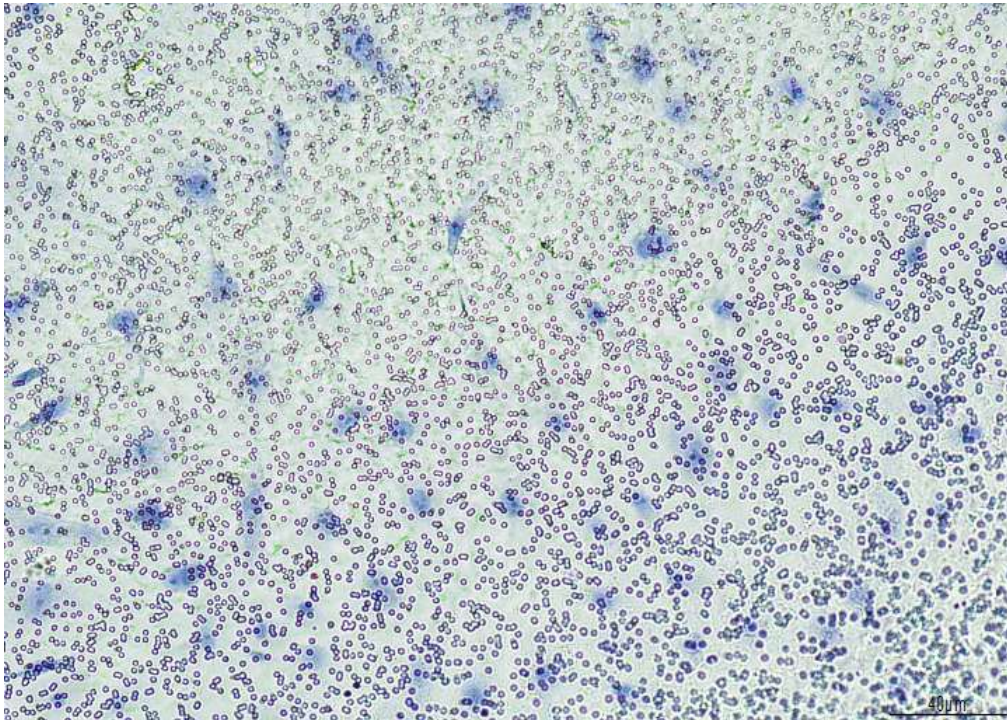
n= 1 insert only, n=3 endothelial barrier (mean  $\pm$  SEM)

0.22mg/mL Evans blue in endothelial medium

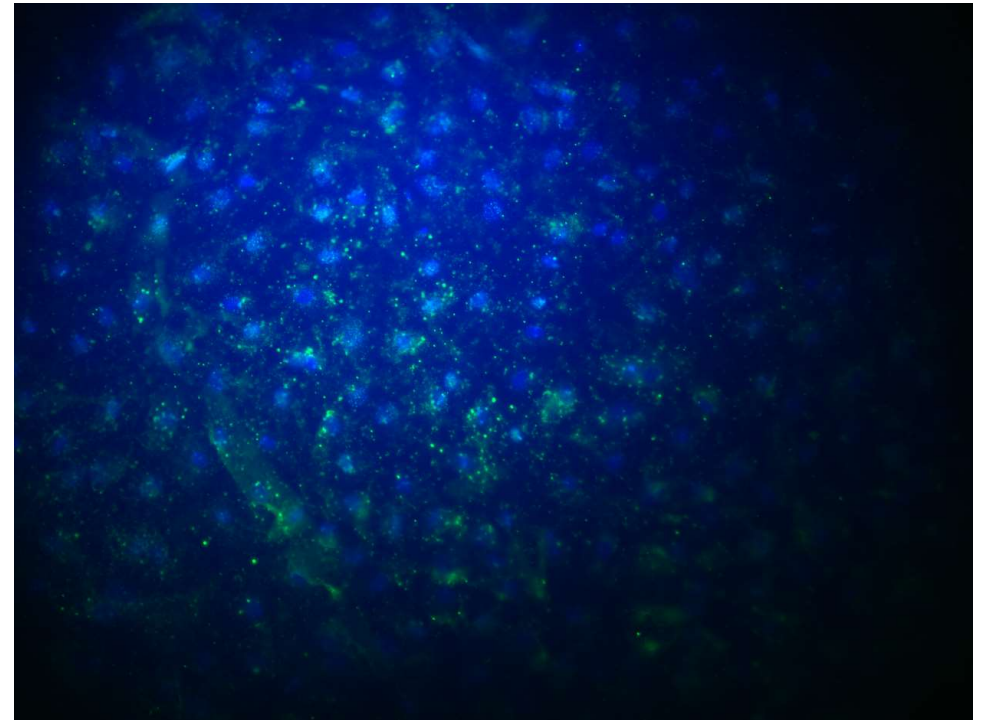


# Separation of compartments via endothelial barrier

HUVEC nuclei stained with haematoxylin on the underside of a 12-well insert

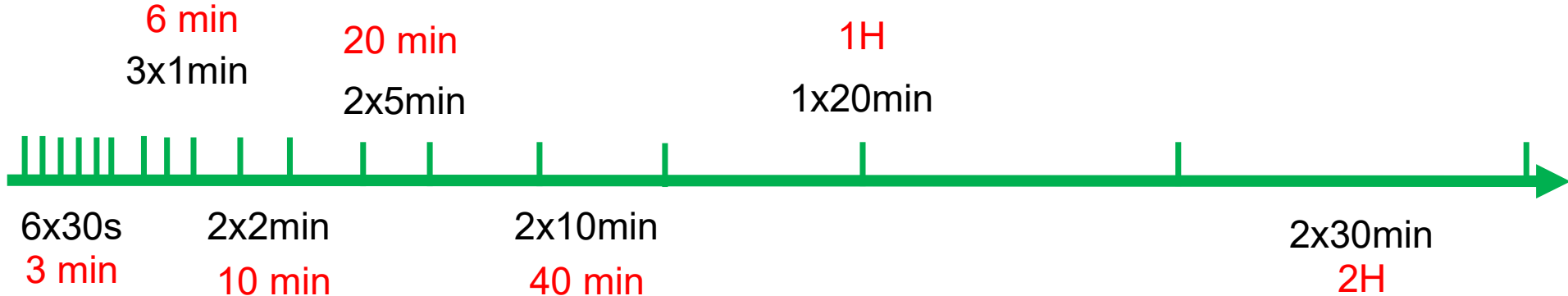


HUVECs stained (badly) with DAPI and CD31 on the underside of a 12-well insert



# Kinetic studies

## Docetaxel



## [<sup>18</sup>F]FDG

